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(21) International Application Number: PCT/US92/10391 (22) International Filing Date: 1 December 1992 (01.12.92) (30) Priority data: 91870208.5 13 December 1991 (13.12.91) EP (34) Countries for which the regional or international application was filed: GB et al. (71) Applicant (for all designated States except US): THE PROCTER & GAMBLE COMPANY [US/US]; One Procter & Gamble Plaza, Cincinnati, OH 45202 (US). (72) Inventors; and (75) Inventors/Applicants (for US only): HARDY, Frederick, Edward [GB/GB]; 8 Woodend/Darras Hall/Ponteland, Newcastle Upon Tyne NE20 9ES (GB). WILLEY, Alan, David [GB/GB]; 17 Brandon Grove, Sandyford, Newcastle upon Tyne NE2 1PA (GB). SCIALLA, Stefano [IT/IT]; Viale dei Caduti nella Guerra di Liberazione, 131, I-00128 Rome (IT).		(74) Agents: REED, T., David et al.; The Procter & Gamble Company, 5299 Spring Grove Avenue, Cincinnati, OH 45202 (US). (81) Designated States: CA, FI, JP, NO, US, European patent (AT, BE, CH, DE, DK, ES, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE). Published <i>With international search report.</i>
(54) Title: ENCAPSULATION OF LIQUIDS IN MICROORGANISMS (57) Abstract Yeast or other microorganism cells for use in the encapsulation of liquids (e.g. liquid bleach activators for use in laundry detergent compositions) are deodorised by treatment with a peroxygen bleach, e.g. hydrogen peroxide.		

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ENCAPSULATION OF LIQUIDS IN MICROORGANISMS

Field of the Invention

The present invention relates to a method of reducing the
5 odour of micro-organism cells. The invention also relates
to the use of the resultant deodorised cells in a process
for encapsulating a material, in which micro-organism cells
are contacted with the said material, which material is in
liquid form, whereby the said material is absorbed through
10 the micro-organism cell wall and retained within the micro-
organism cells. The invention also relates to liquids that
have been encapsulated in that manner.

Background to the Invention

The encapsulation of materials within micro-organism cells
15 is well known. In EP-B-0,085,805 (Dunlop Limited), a method
of encapsulation is described in which the micro-organism is
contacted with an organic lipid-extending substance that is
a solvent, or a micro-dispersing medium, for the material to
be encapsulated, and simultaneously and/or subsequently the
20 micro-organism is contacted with the material to be
encapsulated, said material being employed as a solution or
micro-dispersion in the organic lipid-extending substance,
or in a further organic lipid-extending substance or in a
liquid that is miscible with the first-mentioned lipid-
25 extending substance, whereby both the organic lipid-
extending substance and the material to be encapsulated are
taken into and retained passively within the micro-organism.
Suitable micro-organisms include yeasts and suitable lipid-
extending substances include aliphatic alcohols, esters,
30 aromatic hydrocarbons and hydrogenated aromatic
hydrocarbons. An example of a material that can be
encapsulated is a leuco dye suitable for use in "carbonless"
copy paper. A stated advantage of the method described in
that European patent is that, in contrast to certain earlier
35 proposals (cf. US-A-4,001,480 and FR-A-2,179,528), use may
be made of micro-organisms having a natural lipid content of
less than 40 percent by weight, without the need to employ
a plasmolyser.

EP-A-0,242,135 (AD2 Limited) discloses a method of producing an encapsulated material by contacting the material in liquid form with a grown intact micro-organism having a microbial lipid content of less than 40 percent by weight. The encapsulatable material must be capable of diffusing into the microbial cell without causing total lysis thereof and the treatment of the micro-organism is carried out in the absence of an organic lipid-extending substance as solvent or microdispersant for the encapsulatable material and in the absence of a plasmolyser. The material is absorbed by the micro-organism - typically a yeast - by diffusion across the microbial cell wall and is passively retained within the micro-organism. A wide variety of encapsulatable materials are mentioned, including essential oils used as flavours or fragrances, leuco dyes, vitamins, detergents such as lauryl ether sulfate, food colourants, and pesticides and the like.

EP-A-0,414,282 (Quest International) discloses bleach compositions, including laundry detergents, laundry bleaches and dishwashing or scouring products, that contain a perfume whereas EP-A-0,414,283 (Quest International) discloses fabric-softening compositions that contain a perfume. In both cases, the perfume is encapsulated in micro-organism cells according to the conventional methods, as described in US-A-4,001,480 or EP-A-0,242,135.

A problem that arises when using micro-organism cells for encapsulation purposes is that they may have a disagreeable odour and possibly also an unpleasing colour and may therefore diminish the acceptability of the encapsulated products, or compositions containing them, to consumers.

Summary of the Invention

It has now been found that micro-organism cells may be at least partially deodorised by treatment with a peroxygen bleach whilst leaving the cells at least largely intact and hence suitable for encapsulation purposes.

The present invention, in one of its aspects, accordingly provides a method of reducing the odour of micro-organism

cells, characterised in that the said cells are treated with a peroxygen bleach.

The invention also provides, in another of its aspects, a process for encapsulating a material in which micro-organism
5 cells are contacted with the said material, which material is in liquid form, whereby the said material is absorbed through the micro-organism cell wall and retained within the micro-organism cells, characterised in that the micro-organism cells are also treated with a peroxygen bleach.

10

Description of Exemplary Embodiments

Although bacteria and algae may be suitable, preferred micro-organisms are fungi, especially filamentous fungi, e.g. *Aspergillus niger*, and more especially the yeasts.

15 Examples of yeasts which may be used in the present invention are *Lipomyces* species, such as *L. lipofer* and *L. starkeyi*, *Trichosporon* species, such as *T. pullulans* and *T. cutaneum*, *Candida* species such as *C. curvata* and *C. utilis*, *Kluyveromyces fragilis* and *Saccharomyces cerevisiae*.

20 Usually, use will be made of micro-organism cells in grown form, i.e. that have been harvested from a culture medium. The cells should be intact, that is to say not have undergone any significant lysis, and are preferably of large size, typically with an average diameter for the cell
25 of from 5 to 20 μm . It is also desirable that the cells should not undergo lysis before or during the encapsulation step.

A micro-organism will normally be chosen that under the conditions of the intended use will disintegrate or undergo
30 sufficient disruption or permit diffusion of its contents so that the encapsulated liquid will be released at the appropriate point of application.

In accordance with this invention, the micro-organism cells are treated with a peroxygen bleach, for example peracids
35 (including so-called 'low-activity' peracids), such as peroxymonosulfuric acid, m-chloroperbenzoic acid, diperoxyphthalic acid and monoperphthalic acid, and their derivatives, e.g. salts such as potassium monopersulfate

("oxone") or magnesium monoperoxyphthalate ("H48"). Compounds that release hydrogen peroxide when dissolved in water come into consideration, especially those that are commercially available; these includes such salts as
5 perborates, notably sodium perborate, and such adducts as percarbonates. At present, however, the preferred peroxygen bleach is hydrogen peroxide (H_2O_2).

Although the applicant does not wish to be bound by any theory, it is believed that the peroxygen bleach attacks the
10 amine centres in the micro-organism. Generally, the treatment with the peroxygen bleach should be carried out under conditions that achieve a significant reduction in, or even complete removal of, the characteristic micro-organism odour and/or colour without being so severe as to cause any
15 significant lysis or disruption of the cell walls (which would impair the effectiveness of the micro-organism as an encapsulation material). Generally, the micro-organism is treated with an aqueous solution of the peroxygen bleach, especially one containing the peroxygen bleach at a
20 concentration of from 0.01 to 10%, more preferably from 0.02 to 5% and most preferably from 0.1 to 2%, w/v. Relative to the weight of micro-organism, the amount of peroxygen bleach is usually 0.02 to 100%, preferably 0.04 to 50% and more preferably 0.1 to 20% by weight. The solution of peroxygen
25 bleach is preferably prepared in deionised water.

The peroxygen bleach-containing treatment solution is preferably alkaline and will typically contain an alkali - usually an alkali metal or alkaline earth metal hydroxide or carbonate, preferably sodium hydroxide - at a concentration
30 from 0 to 2.0 M, preferably from 0.01 to 1.0 M, more preferably from 0.05 to 0.5 M. Simply buffering the solution at high pH may also be considered.

It has also proved advantageous for the peroxygen bleach-containing solution to contain sodium silicate, preferably
35 in an amount from 20 to 60g/l, typically from 30 to 50g/l. The sodium silicate is useful as a filtering aid, is a source of alkalinity, acts as a defoaming agent and provides some control over heavy metals that might decompose the

peroxygen bleach. Of course, other silicates, e.g. other alkali metal silicates, come into consideration for use with or instead of the sodium silicate, as do filtration aids, such as Kieselguhr or Celite, and chelating agents, such as phosphonates, ethylenediamine tetraacetic acid (EDTA) and sodium tripolyphosphate (STP).

Normally, up to 250g, typically 50 to 150g, of micro-organism is employed per litre of the peroxygen bleach-containing solution. The treatment is conveniently effected by suspending the micro-organism in the treating solution and gently stirring the suspension. Suitable durations for the treatment and suitable temperatures at which it may be carried out can be determined by simple trials; in general, it has been found adequate for the suspension to be stirred for from 5 minutes to 4 hours, preferably from 30 minutes to 2 hours and typically for about 1 hour, at a temperature of from 0° to 100°C, preferably from 10° to 50°C and typically at room temperature. The treated micro-organism is then separated from the treating solution by any convenient method, e.g. centrifugation, and is generally dried, e.g. by freeze drying, before further use.

Although the primary purpose in treating the micro-organism cells with the peroxygen bleach is to reduce or eliminate any odour of the micro-organism, the treated micro-organism may also be referred to hereinafter as a "bleached" micro-organism; it has been found that the reduction in odour is often accompanied by a lightening of the colour of the micro-organism cell material.

The treated micro-organism may be employed for the encapsulation of a wide variety of encapsulatable materials using any of the methods known in principle from the prior art, e.g. the method described in US-A-4,001,480, in EP-B-0,085,805 or, preferably, in EP-A-0,242,135 (the teaching in each of which is incorporated herein by reference).

The material to be encapsulated should be in liquid form under the conditions at which encapsulation is carried out. Materials that are not themselves liquid under those conditions may be used in the form of a solution or micro

dispersion in a suitable solvent or dispersant, usually a solvent that is immiscible with lipid that may be present in the micro-organism. Suitable solvents include $C_1 - C_4$ alcohols, e.g. methanol, ethanol or isopropanol, and the
5 solvent may be removed by evaporation after the encapsulation treatment. It is also possible, and sometimes preferred, to carry out the encapsulation process in the presence of water.

Although it is preferred to treat the micro-organisms with
10 the peroxygen bleach before encapsulation is effected, it is possible in principle to effect such treatment during or even after the encapsulation step. Thus, for example, encapsulation could be effected from a system containing both the peroxygen bleach in aqueous solution and the
15 material to be encapsulated, provided that the material to be encapsulated were compatible with that bleach.

The present invention is particularly advantageous in the encapsulation of bleach activators used in cleaning compositions, for example heavy duty or general purpose
20 laundry detergent compositions, bleaching compositions, dishwashing compositions and hard-surface or other cleaning products.

Such bleach activators are commonly susceptible to attack by moisture, leading to hydrolysis or premature
25 perhydrolysis, the products of which are liable to damage other ingredients in the cleaning composition. Suitable bleach activators are disclosed in U.S. Patents No. 4,179,390 (Spadini et al.), No. 4,412,934 (Chung et al.) and No. 4,915,854 (Mao et al.)

30 The present invention is illustrated in and by the following examples.

Example 1

(a) Yeast Treatment

100g of baker's yeast (*Saccharomyces cerevisiae*) were
35 suspended in one litre of a 0.2 molar solution of sodium hydroxide in water containing 40g of sodium silicate. Hydrogen peroxide was added until its concentration reached 1% w/v and the resultant suspension was then gently stirred

for one hour at room temperature. The yeast was then removed by centrifugation and freeze dried.

(b) Encapsulation

One part by weight of the bleached yeast obtained according to the above-described treatment (a) was suspended in three parts of water and stirred at 60°C for one hour. 0.6 parts of the material to be encapsulated, namely acetyl triethyl citrate, was then added and the suspension was stirred for 6 hours at 45°C. The yeast cells were then removed by centrifugation and freeze dried. The resultant yeast-encapsulated acetyl triethyl citrate was suitable for incorporation into detergent formulations as a bleach activator.

Example 2

A number of samples of bleached yeast micro-capsules were prepared using the process described above in Example 1 (a) but with certain variations in the yeast concentration, alkalinity, the presence or absence of sodium silicate, and the concentration of hydrogen peroxide. The samples were assessed for yeast odour and for the amount of foaming produced during the hydrogen peroxide treatment. The results are summarised in the following table (in which sample 5 indicates the sample obtained by the process according to Example 1(a)).

Table 1

Sample	Yeast Conc. (g/l)	Alkalinity (M)	Sodium Silicate (+/-)	H ₂ O ₂ (%)	Yeast Odour (1-5)	Foaming (1-5)
1	250	-	-	10	1	5
2	100	-	-	1	1	5
3	100	2	+	10	1	1
4	100	1	+	1	1	1
5	100	0.2	+	1	1	2
6	100	0.05	+	1	3	5

Key

Odour

1 = Best, i.e. little or no odour
5 = Worst, i.e. odour approximately equal to starting material.

5 Foaming

1 = Best, i.e. little foaming
5 = Worst, i.e. excessive foaming

10

For the yeast cells to be useful for encapsulation purposes, the cell membrane must be intact. Microscopy showed that this was only the case for Samples 5 and 6. Sample 5 was preferred to Sample 6, in that it exhibited a lower odour and lower foaming.

15 Example 3

Bleached yeast micro-capsules containing the liquid bleach activator, acetyl triethyl citrate, which micro capsules had been prepared according to the process of Example 1 (b), were blended into a standard detergent composition and stored in sealed cartons under stressed storage conditions (32°C and 80% humidity). For comparison purposes, a similar composition was prepared containing the liquid bleach activator encapsulated in unbleached yeast micro-capsules prepared according to the prior art (EP-A-0,242,135). A further comparison composition was prepared containing, instead of the encapsulated liquid bleach activator, a conventional activator, tetraacetyl ethylene diamine (TAED), in particulate form. The comparison compositions were stored in sealed cartons under the stressed storage conditions specified above.

The compositions were sampled after certain periods of time, the samples being analyzed in order to determine how much of the bleach activator (acetyl triethyl citrate or TAED, as the case may be) remained (expressed as a percentage of the original content). Specifically, the analysis was effected by dissolving the sample, analyzing for peracid and comparing that result with the expected result for 100% active. The results are shown in the following table.

Table 2

	<u>Time</u> <u>Weeks</u>	<u>Unbleached</u> <u>% remaining</u>	<u>Bleached</u> <u>% remaining</u>	<u>TAED</u> <u>% remaining</u>
5	0	100	100	100
	2	81	76	98
	5	67	(29)	67
	8	27	40	61
10	<hr/>			

The 5-week result for the bleached capsules is anomalous and is thought to be due to poor dispersion of the product; in particular, caking, which is a problem commonly experienced when using such stressed storage conditions, tends to render dissolution of the product difficult. Overall, the performance of the bleached capsules in preserving the activity of the bleach activator was deemed comparable to the effectiveness of the conventional, unbleached capsules.

It will of course be understood that the present invention has been described above purely by way of example and that modifications of detail can be made within the scope of the invention.

CLAIMS

1. A method of reducing the odour of micro-organism cells, characterised in that the said cells are treated with a peroxygen bleach.
- 5 2. A method according to claim 1, wherein the said cells are treated in an aqueous medium containing the peroxygen bleach at a concentration of from 0.01 to 10% w/v.
3. A method according to claim 1 or 2, wherein the said cells are treated with the peroxygen bleach under alkaline
10 conditions.
4. A method according to claim 3, wherein the treatment of the said cells with the peroxygen bleach is carried out in the presence of sodium hydroxide at a concentration of up to 2 molar.
- 15 5. A method according to any of claims 1 to 4, wherein the treatment of the said cells with the peroxygen bleach is carried out in the presence of sodium silicate.
6. A method according to any of claims 1 to 5, wherein from 50 to 250g of micro-organism cells are treated per
20 litre of treatment medium comprising the peroxygen bleach.
7. A method according to any of claims 1 to 6, wherein the peroxygen bleach is used in an amount of from 0.02 to 100% by weight, relative to the weight of micro-organism cells.
8. A method according to any of claims 1 to 7, wherein the
25 peroxygen bleach is hydrogen peroxide.
9. A method according to any of claims 1 to 8, wherein the micro-organism cells are yeast cells.
10. A method according to claim 9, wherein the yeast is selected from *Saccharomyces cerevisiae*, *Kluyveromyces*
30 *fragilis* and *Candida utilis*.
11. A process for encapsulating a material in which micro-organism cells are contacted with the said material, which material is in liquid form, whereby the material is absorbed through the micro-organism cell wall and retained within the
35 micro-organism cells, characterised in that the micro-organism cells are also treated with a peroxygen bleach.
12. A process according to claim 11, wherein the cells are treated with the peroxygen bleach using a method as

defined in any of claims 2 to 10.

13. A process according to claim 11 or 12, wherein the cells are treated with the peroxygen bleach prior to contact with the material to be encapsulated.

5 14. A process according to claim 11, 12 or 13, wherein the material to be encapsulated comprises a bleach activator suitable for use in laundry detergent compositions.

15. Micro-organism, e.g. yeast, cells that are substantially intact and that have been treated with a
10 peroxygen bleach, e.g. hydrogen peroxide.

16. A liquid that is encapsulated within micro-organism, e.g. yeast, cells, characterised in that the yeast cells have been also treated with a peroxygen bleach, e.g. hydrogen peroxide.

15

INTERNATIONAL SEARCH REPORT

PCT/US92/10391

A. CLASSIFICATION OF SUBJECT MATTER

IPC(5) :B01J.13/02, 13/04; C11D 3/395; C12N 1/16, 1/18

US CL :252/95, 186.28, 186.38, 186.41; 264/4.1; 428/402.2; 435/255, 256

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

U.S. : 252/95, 186.28, 186.38, 186.41; 264/4.1; 428/402.2; 435/255, 256

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
<u>X</u> Y	US, A, 1,056,540 (Hentschel) 18 March 1913, See page 1, lines 30-46.	<u>1,2,6.-10,15</u> 3-5,11-14,16
<u>X</u> Y	US, A, 2,031,668 (Reich) 25 February 1936, See page 2, lines 9-43.	<u>1,2,6-10,15</u> 3-5,11-14,16
Y	US, A, 3,951,594 (Smolens) 20 April 1976, See column 2, lines 18-36; and table 1.	3-5
<u>X</u> Y	US, A, 4,001,480 (Shank) 04 January 1977, See column 1, lines 46-51 and 63-68; column 2, lines 16-51; column 3, lines 18-40; and Ex. IV.	<u>11-13,16</u> 14

☒ Further documents are listed in the continuation of Box C. ☐ See patent family annex.

* Special categories of cited documents:	*T	later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
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25 FEBRUARY 1993

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C (Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	US, A, 4,025,453 (Kravetz et al.) 24 May 1977, See column 4, line 63 - column 5, line 44.	14
A	US, A, 3,925,234 (Hachmann et al.) 09 December 1975.	11-14,16
A	EP, A, 0,242,135 (Pannell) 21 October 1987.	1-16

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